Contents lists available at SciVerse ScienceDirect

Journal of Microbiological Methods

journal homepage: www.elsevier.com/locate/jmicmeth

Time-to-positivity-based discrimination between *Enterobacteriaceae*, *Pseudomonas aeruginosa* and strictly anaerobic Gram-negative bacilli in aerobic and anaerobic blood culture vials

Gilles Defrance ^a, Gabriel Birgand ^{b,*}, Etienne Ruppé ^{a,c}, Morgane Billard ^a, Raymond Ruimy ^{a,c}, Christine Bonnal ^b, Antoine Andremont ^{a,c}, Laurence Armand-Lefèvre ^{a,c}

^a Bacteriology Laboratory, Bichat-Claude Bernard Hospital, Assistance Publique - Hôpitaux de Paris (AP-HP), 46 rue Henri Huchard, 75877 Paris Cedex 18, France ^b Infection Control Unit, Bichat-Claude Bernard Hospital, Assistance Publique - Hôpitaux de Paris (AP-HP), 46 rue Henri Huchard, 75877 Paris Cedex 18, France ^c EA 3964 University Paris 7, Denis Diderot, 46 rue Henri Huchard, 75877 Paris Cedex 18, France

ARTICLE INFO

Article history: Received 10 January 2013 Received in revised form 15 February 2013 Accepted 15 February 2013 Available online 28 February 2013

Keywords: Time to positivity Bactec Aerobic Anaerobic Vials Discrimination

ABSTRACT

Time-to-positivity (TTP) of first positive blood cultures growing Gram-negative bacilli (GNB) was investigated. When anaerobic vials were positive first, TTP \leq 18 h differentiated *Enterobacteriaceae* from strict anaerobic Gram-negative bacilli (PPV 98.8%). When the aerobic ones were first, TTP \leq 13 h differentiated *Enterobacteriaceae* from *Pseudomonas aeruginosa* and other GNB (PPV 80.8%).

© 2013 Elsevier B.V. All rights reserved.

Gram-negative bacilli (GNB) bloodstream infections are associated with high fatality rates (Leibovici et al., 1998). Poor outcome is associated with inadequate or delayed appropriate antibiotics therapy, and use of empirical broad-spectrum antibiotics is recommended (Leibovici et al., 1998; Garnacho-Montero et al., 2003). However, the struggle against the worldwide rise of bacterial resistance calls for a wise usage of these classes of antibiotics (Boucher et al., 2009). Early discrimination of Enterobacteriaceae, Pseudomonas aeruginosa and anaerobic GNB, which have different susceptibility to antibiotics, would allow a rapid implementation of an antibiotic specific regimen. Commonly used, automated vial incubators for blood cultures record the time elapsed between the introduction of vials into the incubator and positivity detection or time-to-positivity (TTP). In the present study, we estimated TTP diagnostic value to differentiate Enterobacteriaceae, P. aeruginosa and anaerobic GNB in patients with a blood culture vial, either an anaerobic or aerobic one, which was positive first with GNB upon Gram stain examination.

During two years, in our hospital, TTPs of all blood cultures (one Bactec Plus Aerobic/F and one Lytic/10 Anaerobic/F vial; Becton-Dickinson, Le Pont-De-Claix, France), first positive with GNB on direct smear

* Corresponding author at: Unité de Lutte contre les Infections Nosocomiales, GH Bichat-Claude Bernard, 46 rue Henri Huchard, 75877 Paris Cedex 18, France. Tel.: +33 140 256 199; fax: +33 140 258 811.

E-mail address: gbirgand@gmail.com (G. Birgand).

examination, were analyzed. All vials were processed using the Bactec 9240 system (Becton–Dickinson). Vials with more than one microbial species were excluded. *Enterobacteriaceae*, *P. aeruginosa* or strict anaerobic Gram-negative bacilli (SAGNB) were identified using API identification kits (bioMérieux). TTPs, in hours without decimal digit, were aggregated in four time-interval groups with roughly the same number of positive bottles each. Bacterial identification was the gold standard. Clinical data were recorded. Aggregated and continuous TTPs were compared using χ^2 test and Student's *t*-test, respectively. Sensitivity, specificity, positive predictive value (PPV), and positive likelihood ratios (LR+) were calculated for each time interval, and the Receiver Operative Characteristic (ROC) curve was constructed to determine the best TTP cut-off value, as described (Sackett et al., 1991).

Overall, 607 consecutive GNB bloodstream episodes from 596 patients (mean age 67.1 years [range: 1–102]; sex ratio male/female = 0.55) were analyzed. Sources of bacteremia were urinary tract infections (35.3%, n = 214), intra-abdominal infections (16.1%, n = 98), pneumonia (7.4%, n = 45), central venous catheter related infections (4.3%, n = 26), skin and soft tissue infections (3.5%, n = 21), others (7.9%, n = 48), or not documented (25.5%, n = 155). The anaerobic vial was first positive in 394 (64.9%) patients and the aerobic in 213 (35.1%).

In patients with anaerobic vial positive first, *Enterobacteriaceae* were isolated in 343 (87.1%), SAGNB in 49 (12.4%) and other GNB in 2 (0.5%) (1 *Haemophilus influenzae* and 1 *Capnocytophaga* spp.) (Table 1). No



Note



CrossMark

^{0167-7012/\$ -} see front matter © 2013 Elsevier B.V. All rights reserved. http://dx.doi.org/10.1016/j.mimet.2013.02.005

Table 1

Likelihood ratio (LR+) and predictive positive values (PPV) for the presence of strictly anaerobic Gram-negative bacilli (SAGNB) vs *Enterobacteriaceae* and other GNB in anaerobic blood culture vials; and for the presence of *Enterobacteriaceae*, *P. aeruginosa* and other Gram-negative bacilli (GNB) in aerobic blood cultures vials.

Results for aerobic blood o	cultures vials					
Intervals of TTP (h)	Patients with first positive vials (%)		P. aeruginosa		Others	
	<i>P. aeruginosa</i> $(n = 62)$	Others $(n = 151)$	PPV	LR+	PPV	LR+
0-13	14 (22.6)	90 (59.6)	13.5	0.38	86.5	2.6
14–17	19 (30.6)	18 (11.9)	51.4	2.6	48.6	0.3
18–20	14 (22.5)	9 (5.9)	60.9	3.8	39.1	0.3
>20	15 (67.7)	34 (76.2)	30.6	1.1	69.4	0.9
Intervals of TTP (h)	Patients with first positive vials (%)		Enterobacteriaceae		Others	
	Enterobacteriaceae ($n = 124$)	Others $(n = 89)$	PPV	LR+	PPV	LR+
0–13	84 (67.7)	20 (13.2)	80.8	3.0	19.2	0.3
14–17	16 (12.9)	21 (13.9)	43.2	0.5	56.8	1.8
18–20	6 (4.8)	17 (11.2)	26.1	0.3	73.9	4.0
>20	18 (14.5)	31 (20.5)	36.7	0.4	63.3	2.4
Intervals of TTP (h)	Patients with first positive vials (%)		Other GNB		P. aeruginosa & Enterobacteriaceae	
	Other GNB ($n = 27$)	P. aeruginosa & Enterobacteriaceae (n = 186)	PPV	LR+	PPV	LR+
0-13	6 (25.0)	98 (52.7)	5.8	0.4	94.2	2.4
14–17	2 (7.4)	35 (18.8)	5.4	0.4	94.6	2.5
18–20	3 (11.1)	20 (10.7)	13.0	0.4	87.0	1.0
>20	16 (59.3)	33 (17.7)	32.7	3.3	67.3	0.3
Results for anaerobic bloo	d culture vials					
Intervals of TTP (h)	Patients with first positive vials (%)		SAGNB		Others	
	SAGNB $(n = 49)$	Others $(n = 345)$	PPV	LR+	PPV	LR+
0-21	11 (22.4)	344 (99.7)	3.1	0.23	95.5	3.1
22–25	14 (28.6)	1 (0.0)	93.3	98.6	6.7	0.01
26-30	12 (24.5)	0 (0.0)	100.0	$+\infty$	0.0	0.0
>30	12 (24.5)	0(0.0)	100.0	$+\infty$	0.0	0.0

vial grew *P. aeruginosa*. Median TTP was 25 h [interquartile range of 22 h–30 h] for SAGNB and 9 h [8 h–10 h] for *Enterobacteriaceae* (p < 0.001). PPV for the presence of SAGNB increased from 3.1% for 0–21 h TTPs to 93.3% for the 22–25 h TTPs, and LR + increased from 0.2 to 98.6, respectively (Table 1). The PPV for the presence of other GNB (large majority of *Enterobacteriaceae*) was 99.7% when TTP was \leq 21 h. ROC curve analysis showed that a cut-off value of 18 h had the best combined sensitivity (98.8%) and specificity (95.9%) to discriminate *Enterobacteriaceae* from SAGNB; a TTP \leq 18 h was predictive of SAGNB with a PPV of 99.4%; and TTP > 18 h was predictive of SAGNB with a PPV of 92.4%. This result was true whatever the species of *Enterobacteriaceae*.

In patients with aerobic vial positive first, Enterobacteriaceae were isolated in 124 (58.2%), P. aeruginosa in 62 (29.1%), and other GNB in 27 (12.7%) (Acinetobacter spp. n = 8, Campylobacter spp. n = 8, Haemophilus spp. n = 6, Stenotrophomonas maltophilia n = 4, Brucella spp. n = 1, Burkholderia cepacia n = 1 and Flavimonas oryzihabitans n = 1) (Table 1). Median TTP was 11 h [9 h–16 h] for Enterobacteriaceae, 17 h [14 h–20 h] for *P. aeruginosa* (p = NS), and 21 h [15 h–67 h] for other GNB in aerobic vials (p < 0.01 vs *Enterobacteriaceae*). TTP ≤ 13 h was associated with a PPV of 80.8% (LR+: 3.0) for Enterobacteriaceae, whereas TTP between 14 and 20 h was associated with a PPV ranging from 51.4% to 60.9% for P. aeruginosa, with LR + of 2.6 and 3.8, respectively. Other GNB were more likely present when TTP was > 20 h (LR +: 3.3). A TTP cutoff value of 13 h had the best combined sensitivity (67.7%) and specificity (77.5%) to discriminate Enterobacteriaceae from other GNB (including P. aeruginosa). This result was true whatever the species of Enterobacteriacae or strict anaerobes. A decision making algorithm is displayed in Fig. 1.



Fig. 1. Decision making algorithm based on TTP for the distinction of Gram-negative bacilli (GNB) in anaerobic and aerobic blood cultures vials.

We showed that TTP could help in discriminating types of GNB in patients with either an aerobic or anaerobic blood culture vial positive first with GNB upon Gram staining. Previous studies had described that Enterobacteriaceae grew faster than P. aeruginosa, SAGNB or other Gram-negative bacteria (Martinez et al., 2007; Passerini et al., 2009) but the diagnostic value of TTP had not been fully analyzed. We showed here that an 18-hour cut-off discriminated SAGNB from Enterobacteriaceae with a PPV as high as 99.4% in anaerobic vials. TTP analysis was less obvious when aerobic vials were positive first. However, TTP \leq 13 h was associated with a PPV of 80.8% for the presence of Enterobacteriaceae but of only 13.5% for P. aeruginosa, a probability which increased to 60.9% for TTP of 17-20 h. This simple analysis may seem odd when techniques such as fluorescence, hybridization probes (Sogaard et al., 2007), real-time PCR (Cattoir et al.), and mass spectrometry (Kaleta et al.) are emerging for early and accurate diagnostic of bloodstream infections. Unfortunately, these methods are expensive, time consuming, and still not implemented in all laboratories. However, useful TTP analysis has limits. First, cut-off values calculated here with data from one type of automated blood culture system may vary slightly with others. Second, the volume of blood inoculated and the time before loading the vials were not recorded but variations occurred randomly and did not prevent significant patterns to emerge. Third, antibiotics at time of sampling were unknown but TTP has been presented independent of treatments (Liao et al., 2009; Passerini et al., 2009). In this context, our results offer a cost-free, fast and quantitative addition to Gram staining in order to differentiate Enterobacteriaceae from Pseudomonas or SAGNB in blood culture vials with GNB on direct smear examination and to choose the best empiric antibiotic treatments.

Acknowledgments

None.

References

- Boucher, H.W., Talbot, G.H., Bradley, J.S., Edwards, J.E., Gilbert, D., Rice, L.B., et al., 2009. Bad bugs, no drugs: no ESKAPE! An update from the Infectious Diseases Society of America. Clin. Infect. Dis. 48 (1), 1–12.
- Cattoir, V., Gilibert, A., Le Glaunec, J.M., Launay, N., Bait-Merabet, L., Legrand, P., Rapid detection of *Pseudomonas aeruginosa* from positive blood cultures by quantitative PCR. Ann. Clin. Microbiol. Antimicrob. 9, 21.
- Garnacho-Montero, J., Garcia-Garmendia, J.L., Barrero-Almodovar, A., Jimenez-Jimenez, F.J., Perez-Paredes, C., Ortiz-Leyba, C., 2003. Impact of adequate empirical antibiotic therapy on the outcome of patients admitted to the intensive care unit with sepsis. Crit. Care Med. 31 (12), 2742–2751.
- Kaleta, E.J., Clark, A.E., Johnson, D.R., Gamage, D.C., Wysocki, V.H., Cherkaoui, A., et al., Use of PCR coupled with electrospray ionization mass spectrometry for rapid identification of bacterial and yeast bloodstream pathogens from blood culture bottles. J. Clin. Microbiol. 49(1), 345–353.
- Leibovici, L., Shraga, I., Drucker, M., Konigsberger, H., Samra, Z., Pitlik, S.D., 1998. The benefit of appropriate empirical antibiotic treatment in patients with bloodstream infection. J. Intern. Med. 244 (5), 379–386.
- Liao, C.H., Lai, C.C., Hsu, M.S., Huang, Y.T., Chu, F.Y., Hsu, H.S., et al., 2009. Correlation between time to positivity of blood cultures with clinical presentation and outcomes in patients with *Klebsiella pneumoniae* bacteraemia: prospective cohort study. Clin. Microbiol. Infect. 15 (12), 1119–1125.
- Martinez, J.A., Pozo, L., Almela, M., Marco, F., Soriano, A., Lopez, F., et al., 2007. Microbial and clinical determinants of time-to-positivity in patients with bacteraemia. Clin. Microbiol. Infect. 13 (7), 709–716.
- Passerini, R., Riggio, D., Radice, D., Bava, L., Cassatella, C., Salvatici, M., et al., 2009. Interference of antibiotic therapy on blood cultures time-to-positivity: analysis of a 5-year experience in an oncological hospital. Eur. J. Clin. Microbiol. Infect. Dis. 28 (1), 95–98.
- Sackett, D.L., B. H. R., Guyatt, G.H., Tugwell, P., 1991. The interpretation of diagnostic data. In: Little, B.A.C. (Ed.), Clinical Epidemiology: A Basic Science for Clinical Medicine. Little Brown, Boston, MA, pp. 69–152.
- Sogaard, M., Hansen, D.S., Fiandaca, M.J., Stender, H., Schonheyder, H.C., 2007. Peptide nucleic acid fluorescence in situ hybridization for rapid detection of *Klebsiella pneumoniae* from positive blood cultures. J. Med. Microbiol. 56 (Pt 7), 914–917.